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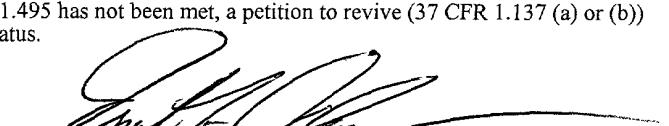
INTERNATIONAL APPLICATION NO. <i>PCT</i> PCT/NZ99/00214	INTERNATIONAL FILING DATE December 10, 1999	PRIORITY DATE CLAIMED January 29, 1999
TITLE OF INVENTION <i>JUL 26 2001</i> TRANSFORMATION AND REGENERATION OF <i>ALLIUM</i> PLANTS		
APPLICANT(S) FOR DO/EO/US Carolyn Elizabeth LISTER and Colin Charles EADY		

Applicant(s) herewith submits to the United States Designated/Elected Office (DO/EO/US) the following items and other information:

1. This is a **FIRST** submission of items concerning a filing under 35 U.S.C. 371.
2. This is a **SECOND** or **SUBSEQUENT** submission of items concerning a filing under 35 U.S.C. 371.
3. This is an express request to begin national examination procedures (35 U.S.C. 371(f)). The submission must include items (5), (6), (9) and (21) indicated below.
4. The US has been elected by the expiration of 19 months from the priority date (Article 31).
5. A copy of the International Application as filed (35 U.S.C. 371 (c)(2)).
 - a. is attached hereto (required only if not communicated by the International Bureau).
 - b. has been communicated by the International Bureau.
 - c. is not required, as the application was filed with the United States Receiving Office (RO/US).
6. An English language translation of the International Application as filed (35 U.S.C. 371 (c)(2)).
 - a. is attached hereto.
 - b. has been previously submitted under 35 U.S.C. 154 (d)(4).
7. Amendments to the claims of the International Application under PCT Article 19 (35 U.S.C. 371 (c)(3)).
 - a. are attached hereto (required only if not communicated by the International Bureau).
 - b. have been communicated by the International Bureau.
 - c. have not been made; however, the time limit for making such amendments has NOT expired.
 - d. have not been made and will not be made.
8. An English language translation of the amendments to the claims under PCT Article 19 (35 U.S.C. 371 (c)(3)).
9. An oath or declaration of the inventor(s) (35 U.S.C. 371 (c)(4)).
10. A copy of the annexes of the International Preliminary Examination Report under PCT Article 36 (35 U.S.C. 371 (c)(5)).

Items 11 to 20 below concern document(s) or information included:

11. Information Disclosure Statement under 37 CFR 1.97 and 1.98.
12. An assignment document for recording. A separate cover sheet in compliance with 37 CFR 3.28 and 3.31 is included.
13. A **FIRST** preliminary amendment.
14. A **SECOND** or **SUBSEQUENT** preliminary amendment.
15. A Substitute specification.
16. A change of power of attorney and/or address letter.
17. A computer-readable form of the sequence listing in accordance with PCT Rule 13ter.2 and 35 U.S.C. 1.821-1.825.
18. A second copy of the published international application under 35 U.S.C. 154 (d)(4).
19. A second copy of the English language translation of the international application 35 U.S.C. 154 (d)(4).
20. Other items or information:
 - a. Copy of cover page of International Publication No. WO 00/44919
 - b. Copy of Notification of Missing Requirements.
 - c.

U.S. APPLICATION NO. (If known, see 37 CFR 1.5) 09/890064		INTERNATIONAL APPLICATION NO. PCT/NZ99/00214	ATTORNEY'S DOCKET NUMBER 01288.0016
21. <input checked="" type="checkbox"/> The following fees are submitted:		CALCULATIONS PTO USE ONLY	
BASIC NATIONAL FEE (37 CFR 1.492 (a) (1) - (5)):			
Neither international preliminary examination fee (37 CFR 1.482) nor international search fee (37 CFR 1.445(a)(2)) paid to USPTO and International Search Report not prepared by the EPO or JPO		\$1000.00	
International preliminary examination fee (37 CFR 1.482) not paid to USPTO but International Search Report prepared by the EPO or JPO		\$860.00	
International preliminary examination fee (37 CFR 1.482) not paid to USPTO but International Search fee (37 CFR 1.445(a)(2)) paid to USPTO		\$710.00	
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Surcharge of \$130.00 for furnishing the oath or declaration later than months from the earliest claimed priority date (37 CFR 1.492 (e)).		<input type="checkbox"/> 20 <input type="checkbox"/> 30	\$
CLAIMS	NUMBER FILED	NUMBER EXTRA	RATE
Total Claims	16	- 20 =	x \$18.00
Independent Claims	2	-3 =	x \$80.00
MULTIPLE DEPENDENT CLAIM(S) (if applicable)		+\$270.00 \$ 270.00	
TOTAL OF THE ABOVE CALCULATIONS =		\$1,270.00	
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SUBTOTAL =		\$ 635.00	
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TOTAL NATIONAL FEE =		\$ 635.00	
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TOTAL FEES ENCLOSED =		\$ 635.00	
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<p>a. <input checked="" type="checkbox"/> A check in the amount of \$ <u>635.00</u> to cover the above fees is enclosed.</p> <p>b. <input type="checkbox"/> Please charge my Deposit Account No. _____ in the amount of \$ _____ to cover the above fees. A duplicate copy of this sheet is enclosed.</p> <p>c. <input checked="" type="checkbox"/> The Commissioner is hereby authorized to charge any additional fees which may be required, or credit any overpayment to Deposit Account No. <u>06-0916</u>. A duplicate copy of this sheet is enclosed.</p> <p>d. <input type="checkbox"/> Fees are to be charged to a credit card. WARNING: Information on this form may become public. Credit card information should not be included on this form. Provide credit card information and authorization on PTO-2038.</p>			
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 SIGNATURE <u>Ernest F. Chapman</u> Reg. No. 25,961 NAME/REGISTRATION NO.			
DATED: July 26, 2001			

AMT 34 AMDT

Transformation and regeneration of *Allium* plants**Field of Invention**

5 The invention relates to a method of transforming plants of the *Allium* family and more particularly to the transformation of onion plants. The invention also relates to the transformed plants.

10 **Background of the Invention**

There are no published protocols for the transformation and regeneration of *Allium* species. The *Allium* crop species are probably the most economically important vegetable species for which transformation technology is unavailable. For other major vegetable crops, confirmed transformation systems have been produced.

15

Initially, many monocotyledons were thought to be unsusceptible to *Agrobacterium*-mediated transformation. The development of direct gene transfer techniques soon led to bombardment being the favoured method of monocotyledon transformation.

20 However, direct gene transfer is not without its problems. Often, low transformation frequencies and a high frequency of unusual integration patterns has been observed in transgenic plants. Recently, *Agrobacterium*-mediated transformation of monocotyledons has gained favour and many monocotyledonous species (including rice, wheat, barley, maize and sugarcane) have now been transformed using this method. A key component in the success of these systems has been the transfer of

25 DNA to callus cell types (usually derived from the pre culture of embryo tissue) followed by regeneration from these callus cells using precise post transformation selection protocols. Transformation of *Allium* callus is not useful as regeneration from callus is extremely difficult.

30 Recently, Haseloff (1997) has modified the *gfp* gene to enhance its use as a transgenic marker gene in viable plant systems. Green fluorescent protein (GFP) enables researchers to follow precisely the fate of any cells expressing this gene and

so optimise post transformation cell survival. Such a system has been useful in the development of the onion transformation protocol reported here.

As monocotyledons, the *Allium* species were predisposed to be recalcitrant to 5 transformation. Onions (*Allium cepa L*) are a crop with diverse environmental requirements. It has, therefore, been relatively understudied with respect to the application of biotechnology. There are only a few reports of DNA delivery to *Alliums* (Klein 1987; Dommissé et al. 1990; Eady et al. 1996; Barandiaran et al. 1998). Three workers used direct gene transfer whilst Dommissé et al.(1990) demonstrated that 10 *Agrobacterium*-mediated transformation may be possible. Recently some reports of regeneration protocols for *Alliums* that are appropriate for transformation study have become available (Hong and Deberg 1995; Xue et al. 1997; Eady et al. 1998; Saker 1998). Only one report exists on the development of potential selective agents for use 15 in *Allium* transformation (Eady and Lister 1998a).

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Object of the Invention

It is therefore an object of the invention to provide a method for producing transgenic *Allium* plants or to at least provide the public with a useful choice.

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In this specification we report the first repeatable protocol for the production of transgenic *Allium* plants.

Summary of the Invention

The invention provides a method of transforming *Allium* plants.

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Preferably, the invention provides an *Agrobacterium tumefaciens* - mediated transformation method for *Allium* plants.

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In particular, the invention provides a method of transforming plants of the *Allium* genus comprising inoculating an embryo culture of an *Allium* species with an *Agrobacterium tumefaciens* strain containing a suitable vector or plasmid.

(followed by page 2a)

AMT 34 AMOT

2a

In particular, the invention provides a method of transforming plants of the *Allium* genus comprising the following steps:

- 5 (a) delivering previously manipulated DNA into embryo, or embryo derived culture cell types of the *Allium* genus via vector or direct gene transfer;
- (b) selecting transformed plant material;
- (c) culturing and regenerating the transformed plants;

wherein the transformation is carried out without passage through a callus phase.

10

(followed by page 3)

ART 34 AMDT

Preferably embryos are inoculated immediately following their isolation.

5 Preferably the transformed plants are onions (*Allium cepa L*).

Preferably immature embryos are used as the explant source.

Preferably the embryos are transformed using a binary vector and more preferably

10 a binary vector carrying a selectable gene.

The embryos may preferably be transformed with a herbicide selective gene.

Examples include the *bar* gene or *prt* gene encoding resistance to phosphinothricin or genes encoding resistance to glyphosate. However other genes may be used.

15

The embryos could alternatively be transformed with an antibiotic selective gene.

An example is the kanamycin or geneticin resistance gene, *nptII*.

In particular, the invention provides a method of transforming *Allium* using

20 immature embryos as an explant source, including:

- a) isolating immature embryos of the *Allium* plant to be transformed;
- b) inoculating the immature embryos with an *Agrobacterium tumefaciens* strain containing a binary vector;
- c) wounding embryos and infiltrating embryos with Agrobacteria;
- d) transferring embryos to a selective medium;
- e) culturing embryo pieces;
- f) selecting putative transgenic cultures; and
- g) regenerating phenotypically normal plants.

30 The invention also provides transformed *Allium* plants. Preferably the *Allium* plants are transformed using protocols in line with the method of the invention.

(followed by page 3a)

AMENDMENT

3a

Callus: uniform Undifferentiated mass of dividing plant cells (Walden, R. (1988).

In: Genetic transformation in plants. Oxford University Press, ISBN 0-335-15822-

5 6) or a tissue arising from disorganized proliferation of cells either in culture or in nature. As opposed to a culture which consists of growing cells, tissues, plant organs, or whole plants in nutrient medium under aseptic conditions e.g. embryo culture (Plant tissue culture: theory and practice. Ed Bhojwani, S.S and Razdan, M.K. 1983, Elsevier) i.e population of differentiated proliferating cells.

10

Brief Description of the Drawings

Embodiments of the invention are now described, by way of example only, with

(followed by page 4)

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reference to the drawings in which:

Figure 1 shows a) GFP expression in embryo tissue after 5 days of cocultivation (x50). b) GFP expression after 2 weeks (x50). c) GFP sector after 6 weeks culture 5 (x25). d) Independent GFP positive tissue (x5). e) GFP positive onion shoot culture (x5). f) Two GFP negative (left) and two GFP positive (right) roots from independent plants (x10). g) Transgenic onion plant (x0.2).

Figure 2 shows Southern analysis of the *gfp* gene of primary transformants: Bluescript 10 plasmid containing the *gfp* gene (uncut), 1 copy number control (lane 1), 10 copy number control (lane 2), blank (lane 3), non transgenic onion (lane 4), 7 transgenic onion plants (lanes 5-11), bluescript plasmid containing the *gfp* gene (uncut), 1 copy number control (lane 12), 10 copy number control (lane 13), blank (lane 14), non transgenic onion (lane 15), 6 transgenic onion plants (lanes 16-21).

Figure 3 shows a Southern blot transgenic antisense root alliinase plants probed with 15 the *gfp* gene fragment to indicate the presence of the pBINmgfpERantirroot T-DNA sequence. Lane 1 lambda hindIII marker; lane 2 one copy equivalent control pBINmgfpERantirroot, lane 3 five copy control pBINmgfpERantirroot; lane 4 non transformed onion, lane 5 positive control onion transformed with pBINmgfpER; lane 20 6-10 transgenic plants transformed with pBINmgfpERantirroot (6&7 and 9&10 are separate clones); lane 11 one copy equivalent control pBINmgfpERantirroot, lane 12 five copy control pBINmgfpERantirroot.

Figure 4 shows a Western blot analysis of alliinase levels in the roots of transgenic 25 and non transgenic onion roots. Lane 1 purified root alliinase control; Lane 2-5 alliinase levels from the roots of four transgenic plants transformed with the pBINmgfpERantirroot DNA; Lane 6 alliinase levels from the roots of a typical non transgenic control plant.

Figures 5 and 6 show a Southern blot analysis of HindIII digested DNA from Onion 30 plants transformed with the modified pCambia 3301 T-DNA. Figure 5 probed with *gfp* probe. Figure 6 probed with *bar* gene probe. Lane 1 and 2, 1 and 5 copy number controls of plasmid pBINmgfpER respectively (containing *gfp* gene), Lane 3 non-transgenic onion DNA. Lane 4-6 clones of a transformant selected from

experiment 994, Lane 7-9 clones of a transformant from experiment 9911. Lane 10-12 control transgenic plants containing the *gfp* gene and not the *bar* gene. Lane 13-14, 5 and 10 copy controls of plasmid containing the *bar* gene (lane 11-14 over washed the *gfp* probed blot).

5 **Figure 7** shows a comparison between A, an onion leaf containing the *bar* gene (two on left) and onion leaves without the *bar* gene (four on right) 10 days after painting with 0.5% v/v solution of the herbicide Buster and B, C and D showing an onion plant without the *bar* gene (left) and containing the *bar* gene (right) 0, 3 and 10 days respectively after spraying with 0.05% vv solution of the herbicide Buster.

10

Detailed Description of the Invention

Materials and Methods

Plant material: Field grown, open-pollinated umbels of *Allium cepa* L. were used as a source of immature embryos. Immature embryos were isolated as described by Eady et al. (1998).

Bacterial strain: *Agrobacterium tumefaciens* strain LBA4404 containing the binary vector *pBIN m-gfp-ER* (Haseloff 1997) or binary vectors derived from p Cambia series were used. Cultures were grown to log phase in LB media containing an appropriate antibiotic and then stored at -80°C in 1 ml aliquots containing 15% glycerol. Aliquots were used to inoculate 50 ml overnight cultures. The following morning cultures were replenished with an equal volume of LB containing antibiotic and 100 µM acetosyringone and grown for a further 4 hours. *Agrobacteria* were isolated by 10 minute centrifugation at 4500 rpm and resuspended in an equal volume of P5 (Eady and Lister 1998a) containing 200 µM acetosyringone.

Transformation procedure: Isolated immature embryos were isolated in groups of 20-40, cut into approximately 1mm sections and then transferred into 0.8 ml of *Agrobacteria* and vortexed for 30 seconds. Following this treatment, embryos were placed under vacuum (~ 20 in. Hg) for 30 minutes before blotting dry on sterile Whatman #1 filter paper and then transfer to solid P5 medium (Eady and Lister 1998) (~40 embryos per plate). After 6 days cocultivation, embryo pieces were transferred

to P5 plus 10 mg/l geneticin and 200 mg/l timentin or 5 mg/l of Basta (active ingredient phosphothrinicin) and 200 mg/l of timentin depending on which binary vector was used. These embryo pieces were cultured in the dark under the same conditions as described for the production of secondary embryos (Eady et al. 1998). Cultures 5 were transferred to fresh medium every 2 weeks. After 3-4 transfers, growing material was transferred to P5 plus 25 mg/l geneticin or 5 mg/l Basta depending on the binary vector used and grown for a further 8 weeks. During this time pieces of putative transgenic tissue that were obviously actively growing were transferred to regeneration medium (Eady et al. 1998). Shoot cultures were maintained for 12 weeks 10 and developing shoots were transferred to 1/2MS media (Murashige and Skoog 1962) plus 20 mg/l geneticin or 5 mg/l of Basta as appropriate to induce rooting. Rooted plants were either transferred to 1/2MS plus 120 g/l sucrose to induce bulbing or transferred to soil in the glasshouse(12 h 12-23°C day, 12h 4-16°C night).

15 *Analyses for transformation:* For GFP expression, tissue was examined by observation under a fluorescence microscope (excitation 475 nm, emission 510 nm Haseloff et al.1997). Larger tissues with high levels of expression were observed using hand held "shirt pocket" fluorescent lanterns (Zelco industries inc., 630 So. Columbus Ave, Mt Vernon NY 10551-4445). *NptII* expression was determined by the 20 ability of regenerating plantlets to form roots on 1/2MS containing geneticin, *bar* expression was determined by the ability of regenerating plantlets to form roots on $\frac{1}{2}$ MS containing Basta.

DNA isolation was performed using a nucleon phytopure plant DNA extraction kit 25 (Amersham Lifescience, Buckinghamshire, England). Southern analysis followed the method of Timmerman et al. (1993) and used PCR-amplified probes to confirm the presence of the *gfp*, *nptII* and *bar* genes. Genomic DNA from the onions was digested with *HindIII*, which cuts once in the middle of the T-DNA.

30 *Cytology:* Chromosome counts were made from the root tips of 2 primary transformants and followed the procedure of Grant et al. (1984).

Example 1*Transformation and characterisation of primary transgenic tissue*

After three days of cocultivation, single cells expressing GFP could be observed in tissue transformed with T-DNA containing the mgfpER gene. Attempts to count cells expressing GFP after 5 days were abandoned as the variation within treatments and between embryo pieces was huge, with many embryo pieces showing no fluorescence and some exhibiting hundreds of fluorescing cells (Fig. 1a). In the latter case, distinguishing between multicellular 'stable' transformation events and multiple adjacent single-celled 'transient' events was not possible. Thus, large biases in any measurement of initial transfer could have occurred. As an alternative, treatments were given an initial transfer rating: *** being excellent initial transfer (~20-30% of tissue pieces with > 20 GFP positive cells per plate), ** represented average initial transfer (5-20% of tissue pieces with some GFP positive cells per plate, and * being poor initial transfer (< 5% of tissue pieces with GFP positive cells per plate) (Table 1). Contamination was a problem in many experiments. Often whole experiments (data not shown) had to be abandoned due to contamination, much of which probably arose from infected embryos.

Table 1. Summary of 5 transformation experiments. * - poor, ** - average, *** - excellent initial transfer, see text for details. Numbers in brackets represent the percentage of transformants from uncontaminated embryos (a - represents the stage and treatments which were transferred to the wrong selective media for 4 days).

Expt	Nº. of embryos	% of embryos contaminated	Initial transfer	Nº. of multicellular GFP tissue pieces		Independent plants	Positive Southern#
				4wk	8wk		
1	~400	100	*	-	-	-	-
2	~360	40	***	52(16) a	15(4.6)	2 (0.6)	1 of 1 tested
3	~440	0	***	72 (16)	44 (10)	12 (2.7)	8 of 8 tested

4	~520	60	**	a	11 (2.4)	3 (0.7)	2 of 2 tested
5	~200	100	*	-	-	-	-

After 2 weeks on selection medium, embryo pieces transformed with pBINmgfpER construct were screened for GFP expression and only pieces containing fluorescing cells were maintained (Fig. 1b). The vast majority of fluorescing cells died over the following four weeks. Some fluorescing cells divided into multicellular clusters of up to ~50 before their ability to fluoresce gradually faded. One interpretation of this was that the transformed cells were still reliant on surrounding non-transgenic cells, which died due to selection pressure and could no longer support the transgenic cells. The 10 number of stable transgenic sectors arising from different plates within experiments varied from 0 to 21 and reflected the numerous parameters that affect the onion transformation process. Comparison between experiments was initially possible and ranged from * in contaminated samples to *** in non-contaminated samples. Indeed, lack of good initial transfer was often an early indication of contamination. Eady et 15 al. (1998) and Eady and Lister (1998ab) demonstrated that genotype, condition of the embryo, size of the embryo, cocultivation conditions and selection pressure all affect embryo survival. The combined effects of these parameters and their interaction with the transformation process will, until they can be controlled, continue to make the success of onion transformation susceptible to large variation.

20

Example 2

Regeneration

After 6 weeks of culture, tissue was transferred to a selective medium without timentin. No growth of *Agrobacterium* was observed in any of the cultures grown on 25 this medium. Fluorescing sectors continued to grow on this media and after 2 transfers it was possible to isolate the first sectors free from non-fluorescing cells (Fig. 1c). As sectors became independent (Fig. 1d) they were transferred to regeneration medium. A few sectors still attached to non-fluorescing tissue were also transferred. On regeneration medium transgenic cultures responded in the same way

as non-transgenic, embryo-derived cultures (Eady et al. 1998). Multiple shoots formed on many of the independent transgenic cultures. However, some, particularly the slower growing or more friable dedifferentiated cultures, either failed to regenerate or produced highly hyperhydric shoots that could not be transferred to the glasshouse.

5 Up to 29% of stable sectors produced shoot cultures from which plants could be obtained (Table 1). These responses to regeneration are typical of those seen in non-transformed embryogenic cultures (Eady et al. 1998). Actively growing shoots were transferred to rooting medium containing an appropriate selective agent. In the instances where non-fluorescing cells were also transferred to shoot media some 10 shoots were produced that failed to root on geneticin. These did not fluoresce. All plants that formed actively growing roots on geneticin also fluoresced (Fig. 1f), indicating that in all instances the complete T-DNA was transferred. Fluorescence in the differentiated structures varied, with most fluorescence being seen in root tips. In shoots, strong fluorescence was limited to young shoots (Fig. 1e). However, GFP 15 fluorescence in shoots was usually masked by red autofluorescence from the chlorophyll. The presence of GFP fluorescence in older leaves could sometimes be observed in the stomatal guard cells.

20 The multiple shoot cultures enabled clonal plants from independent transgenic events to be grown. This was particularly important as earlier attempts to exflask putative transgenic plants had failed (Eady and Lister 1998b). In the first successful transformation experiment described here only 4 from 48 transgenic plants transferred to the soil have died. A total of 14 independent transformants have been transferred to the containment glasshouse.

25

Example 3

Analyses of transformants from plants transformed with pBINmgfpER

30 Apart from fluorescence and growth on geneticin, transformation of onion plants was confirmed by Southern analysis, probing with the *gfp* gene (Fig. 2). As *Hind*III cuts the T-DNA only once it was possible to show copy number from the Southern analysis. Ten of the 13 transformants shown have single copies. The other 3 have 2 (lane 8), 3 (lane 18) and multiple copies (lane 7). Lanes 19 and 21 are from clonal shoots and,

as expected, they show the same pattern. *EcoR1* digest and subsequent Southern analysis liberated an expected internal T-DNA fragment of ~ 900 bp.

Chromosome counts in the 2 primary transformants tested showed a diploid (2n = 16)
5 chromosome complement.

Example 4

Evidence that the transgenic onion plants transformed with the pBINmgfpERantirroot contain the antisense root alliinase gene construct.

10 Onion immature embryos were transformed according to the protocol of Eady et al (1999) with the *pBINmgfpER* plasmid (Haseloff 1997) modified to contain the antisense root alliinase gene construct. The BamH1 to Kpn1 fragment of the root alliinase gene was cloned into the BamH1 - Kpn1 sites in the cloning vector pART7.

15 This gave a antisense version of the root alliinase sequence under control of the CaMV35s regulatory element and *ocs* termination sequence in pART7. The *not1* fragment of this modified pART7 (containing the above *CaMV35s* promoter - antisense root alliinase - *ocs* termination) was then cloned into the Hind111 site of *pBINmgfpER*. Digestion of this plasmid (*pBINmgfpERantirroot*) with BamH1 to liberate 20 a 1.6Kb fragment was used to determine presence and orientation of the insert.

PBINmgfpERantirroot was electroporated into *Agrobacterium tumefaciens* strain LBA4404 and grown on kanamycin to select for transformants. Presence of the binary vector was confirmed by plasmid isolation and PCR for the *gfp* gene. LBA4404 (pBINmgfpERantirroot) was used in transformation experiments.

25 Six putative transformants that fluoresced (to indicate the presence of the *gfp* gene) and grew on media containing geneticin (to indicate presence of the *nptII* gene) were obtained from three experiments. Three of these transformants or clones thereof were analysed by Southern Blot analysis for the successful transfer of the T-DNA insert 30 from the binary vector by probing with both *gfp* and *nptII* gene probes. Roots from these plants were also analysed biochemically for root alliinase enzyme activity following the protocol of Clark et al (1998) (Table 2). Western Blots of the desalted protein (0.5 μ g/lane) extracts were probed with an anti-alliinase antibody and visualised

colourmetrically using a goat-antirabbit-alkaline phosphatase system to determine the relative levels of alliinase enzyme in the transgenic plants.

Table 2:

5	Plant	Alliinase activity (U/mg protein)
Non transgenic CLK control (9910)	14.0	
transformant 992.11F1	3.4	
transformant 994.7G1	11.9	
transformant 992.11F2	9.6	
10 transformant 992.9A1	6.3	

Results

1. Southern Analysis

All three plants analysed contained at least one copy of the T-DNA sequence containing the antisense root alliinase DNA sequence (Figure 3) indicating that integration of modified alliinase sequences into *Allium* species had been achieved. Both *nptII* and *gfp* sequences which flanked the antisense alliinase gene on the T-DNA could be deleted indicating successful transfer of the complete T-DNA in all cases.

20 The Western blot of Figure 4 shows the relative amounts of the root alliinase in protein extracts taken from the transgenic and control roots. These extracts were run on a 10% SDS page gel to separate the proteins and then transferred to nitrocellulose paper using standard techniques. This was then incubated with rabbit polyclonal antibodies raised against the purified alliinase (Clark S. A. 1993. Molecular cloning and 25 cDNA encoding alliinase from onion (*Allium cepa* L.), Ph D. thesis, University of Canterbury, Christchurch, New Zealand). These antibodies have been shown to bind specifically to the alliinase protein. Goat anti-rabbit alkaline phosphatase was added to specifically bind this antibody and after washing, the membrane was immersed in NBT (4 nitrotetrazolium chloride) and BCIP (5 bromo 4 chloro 3 indolyl phosphate) for 30 minutes in the dark. Colour develops at the site of the phosphatase in proportion to the amount alliinase present. The Western blot therefore shows the relative amounts of alliinase protein present in the roots of the transgenic and control onion

plants. The control onion plant has the greatest colour development and has the most alliinase per unit of root protein. The intensity relates to the activity of the enzyme shown in the table and indicates that the activity is related to the amount of alliinase protein and not changes in enzyme activity. This is what is expected when using antisense technology to reduce enzyme activity.

5

Example 5

Transformation of onions to confer herbicide resistance.

Onion immature embryos were transformed according to the protocol with the plasmid 10 pCambia3301 modified, using standard cloning techniques, to contain the *mgfpER* reporter gene construct instead of the *gus* reporter gene. This construct, contained within its T-DNA region the *bar* gene encoding resistance to the herbicide phosphinothricin and the visual reporter gene *mgfpER* both under regulatory control of the CaMV35s promoter. In two separate experiments, 994 and 9911 onion 15 immature embryos from cultivar Cron 19 and CLK respectively were transformed with the above construct. In experiment 994 transformants were selected using the visual marker (GFP expression) and growth on herbicide, whilst on P5 media only. In experiment 9911 only selection on herbicide was used to select for transformation. Selection conditions in both experiments consisted of growth on P5 media containing 20 5mg/l of the herbicide Basta (containing phosphinothricin) and 200mg/l of timentin for 4-6 weeks with fortnightly subculture. Following this, cultures were transferred to P5 media plus 5mg/l Basta for a further 4 weeks of culture. Cultures were then transferred to SM4 media for 6 weeks. In experiment 9911 the SM4 media included 5mg/l Basta. Shoots from 9911 were rooted on 1/2MS30 plus 5mg/l Basta. Shoots 25 from experiment 994 were just rooted in 1/2MS30. Once vigorous root growth was established plants were transferred to the glasshouse.

Results

Putative transgenic plants were produced from both experiments 3 from 994 and 4 from 9911. Southern Blot analysis of clones of one transformant from each 30 experiment demonstrated that the *gfp* gene was present in plants from both experiments and that both cultivars could be transformed (Figure 5). When this blot was subsequently reprobed for the presence of the *bar* gene (Figure 6) only the plants

selected solely on the basis of the herbicide resistance were shown to contain the *bar* gene. These plants were then used for herbicide leaf paint assays and subsequently sprayed with recommended field application rates of the herbicide Buster (active ingredient phosphinotricin). Control plants containing no *bar* gene showed noticeable 5 wilt after 3 days and were essentially dead after 10 days following application of the herbicide, whilst the plants that contained the *bar* gene and had been selected on herbicide did not appear to be harmed and grew normally (Figure 7).

Example 6

10 *Demonstration that transformation is independent of cultivar, construct, T-DNA and selective agent.*

Further to examples 1-5, additional transgenic plants containing variations of the antisense alliinase construct (outlined in example 4) have been produced in additional 15 cultivars CRON 12, CRON19 and CRON2. The nature of these plants has been confirmed by GFP expression where appropriate and regeneration on media containing geneticin. A summary of the plants produced in all the examples outlined above is given in table 3 to demonstrate the plasticity of the transformation system. In experiments that were not contaminated transgenic plants have been obtained from 20 all cultivars so far tested.

Table 3 illustrating the different binary vectors, T-DNA, cultivars and selective agents used in the *Allium* transformation protocol outlined and the measures taken thus far to determine the nature of the transformants.

Experiment number	Binary vector used for transformation (T-DNA in brackets)	Cultivar used	Number of independent transformed lines produced	Number of plants produced	GFP +ve	Growth on selective agent (geneticin or Basta)	Southern Analysis and probe used
98/7,8,9	pBIN(mgfpER)	CLK	>10	>100	yes	yes (geneticin)	12/12 tested (gfp)
99/2	pBIN(antiroot-mgfpER)	CRON12	3	>40	yes	yes (geneticin)	5/5 tested (gfp & nptII)
99/3	pBIN(35santibulb-mgfpER)	CRON19	1	12	yes	yes (geneticin)	yet to be tested
99/3	pBIN(35santibulb-mgfpER)	CRON2	3	7	yes	yes (geneticin)	yet to be tested
99/4	pBIN(antiroot-mgfpER)	CRON19	1	3	yes	yes (geneticin)	yet to be tested
99/4	pCambia3301 (modified)	CRON19	1	10	yes	? (phosphinothricin)	3/3 (gfp) tested 0/3 (bar) tested
99/6	pCambia3301 (modified)	CRON12	1	2	yes	? (phosphinothricin)	Yet to be tested
99/6	pBIN(antiroot-mgfpER)	CRON12	1	1	yes	yes (geneticin)	yet to be tested
99/7	pBIN(mgfpER)	CRON19	1	>12	yes	yes (geneticin)	yet to be tested
99/11	pCambia3301 (modified)	CLK	3	>10	?	Yes (phosphinothricin)	3/3 (gfp & bar) tested
99/12	pBIN(bulbpromoterantibulb-mgfpER)	CLK	1	1	yes	yes	yet to be tested

(Basta = phosphinothricin)

Discussion

5 We have developed a repeatable transformation system for onion. The regenerating primary transformants appear to be phenotypically normal. The GFP expression, as a visual selectable marker, enabled post transformation selection conditions to be optimised. The GFP marker has also proved useful in the selection of transgenic plants from other species that are difficult to transform (Vain et al. 1998). Selection conditions have now been established, which enable the identification of transformants solely on their ability to root in selective media (example 5).

10

15 This method of producing transgenic onions is repeatable and efficient. It takes a short time to produce transgenic plants and utilizes techniques have been shown to be cultivar independent (Example 6).

For example, this described process of transformation can be used with any species within the *Allium* and is not limited to onions. Work has shown that the described process of transformation is genotype independent.

5 It is to be understood that the scope of the invention is not limited to the described embodiments and therefore that numerous variations and modifications may be made to these embodiments without departing from the scope of the invention as set out in this specification.

10 **Industrial Applicability**

The invention provides a novel method of transforming plants of the genus *Allium* and in particular onion plants. Also provided are plants transformed by the method. This allows *Allium* crop species which are an economically important vegetable species to 15 be transformed by a variety of genes for improvement of *Allium* crop varieties.

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Claims

1. A method of transforming plants of the *Allium* genus comprising the following steps:
 - 5 (a) delivering previously manipulated DNA into embryo, or embryo derived culture cell types of the *Allium* genus via vector or direct gene transfer;
 - (b) selecting transformed plant material;
 - (c) culturing and regenerating the transformed plants;wherein the transformation is carried out without passage through a callus phase.
- 10 2. A method according to claim 1 wherein the *Allium* genus is transformed with a strain of *Agrobacterium*.
3. A method according to any one of claims 1-2 in which the plants are onions.
4. A method according to any one of claims 1-3 wherein the embryos are 15 transformed with a binary vector.
5. A method according to any one of claims 1-4 in which embryos of an *Allium* species are inoculated immediately following their isolation.
6. A method according to any one of claims 1-5 in which immature embryos are used.
- 20 7. A method of transforming *Allium* using immature embryos as an explant source, including:
 - (a) isolating immature embryos of the *Allium* plant to be transformed;
 - (b) innoculating cultures of the immature embryos with an *Agrobacterium tumefaciens* strain containing a binary vector;
- 25 (c) wounding embryos and infiltrating embryos with agrobacteria;
- (d) transferring embryos to a selective medium;
- (e) culturing embryo pieces;
- (f) selecting putative transgenic cultures; and
- (g) regenerating plants.
- 30 8. A method according to any one of claims 1-7 wherein the plant is transformed with an *Agrobacterium tumefaciens* strain containing a vector which carries a selectable gene.

9. A method according to claim 8 in which the selectable gene is a herbicide resistance gene.

10. A method according to claim 9 in which the herbicide resistance gene is the *bar* gene or a glyphosate resistance gene.

5 11. A method according to claim 8 in which the selectable gene is an antibiotic resistance gene.

12. A method according to claim 11 in which the antibiotic resistance gene is the *nptII* gene.

13. A method according to any one of claims 1-12 wherein the plant is 10 transformed with a modified alliinase gene.

14. A transformed plant produced by the method of any one of claims 1-13.

15. A transformed plant produced by the method of any one of claims 1-9 in which the resulting transformed plant contains a modified gene involved in sulphur pathway assimilation or breakdown and as a result has altered levels 15 of sulphur compounds.

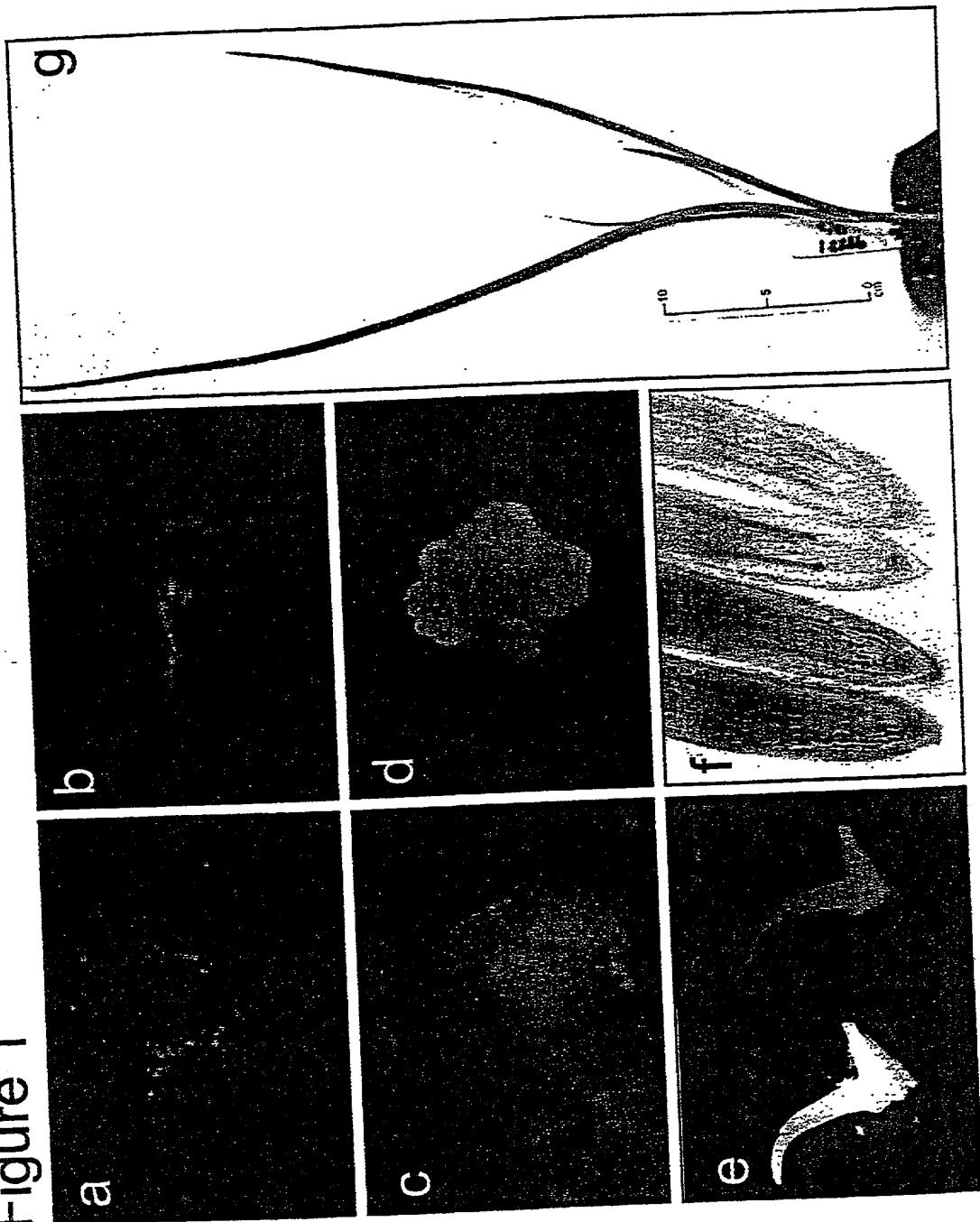
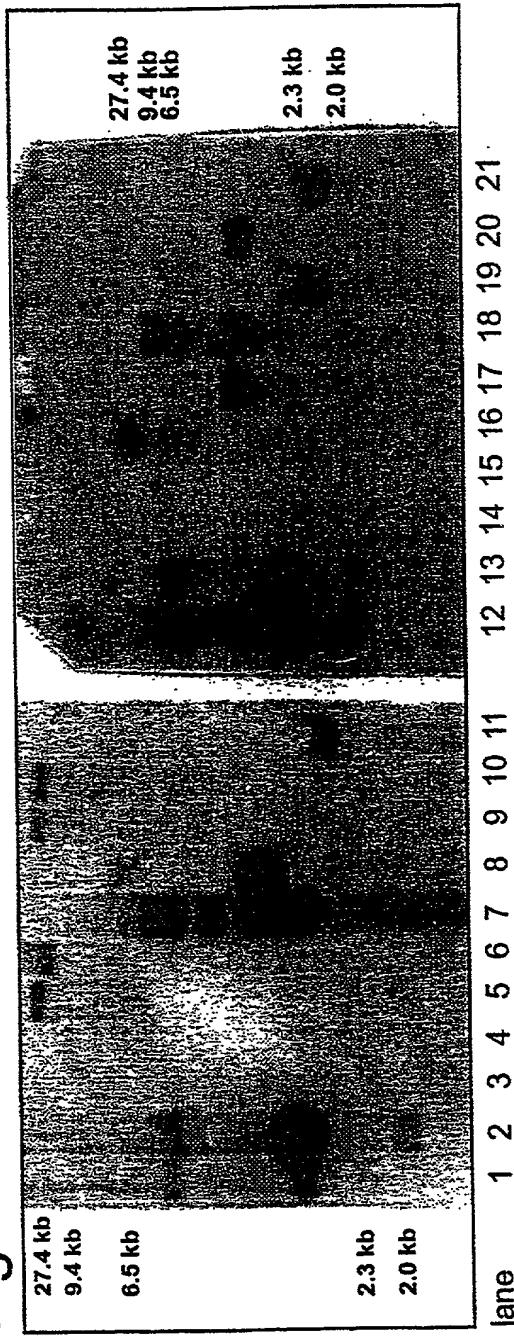


Figure 1

Figure 2



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Figure 3

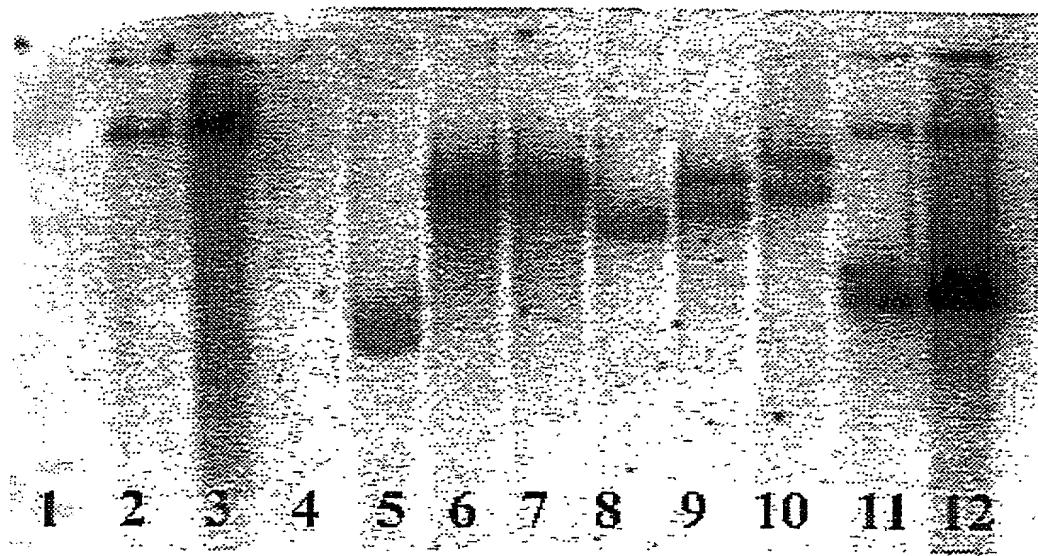
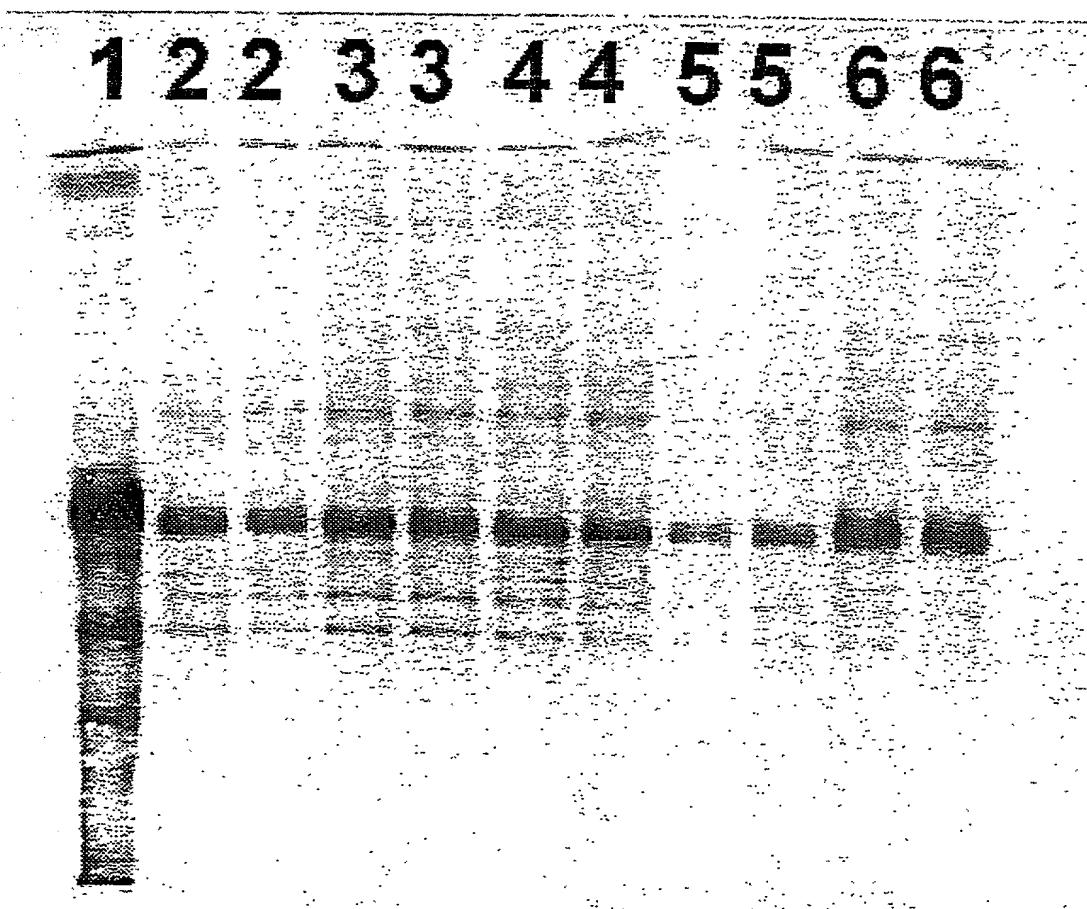


Figure 4



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FIGURE 5

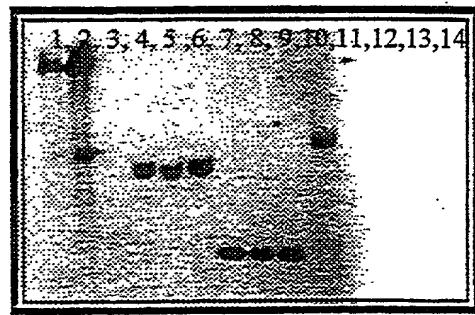
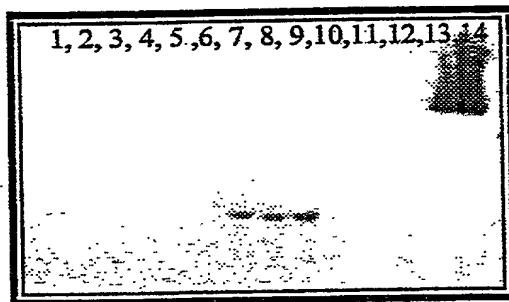
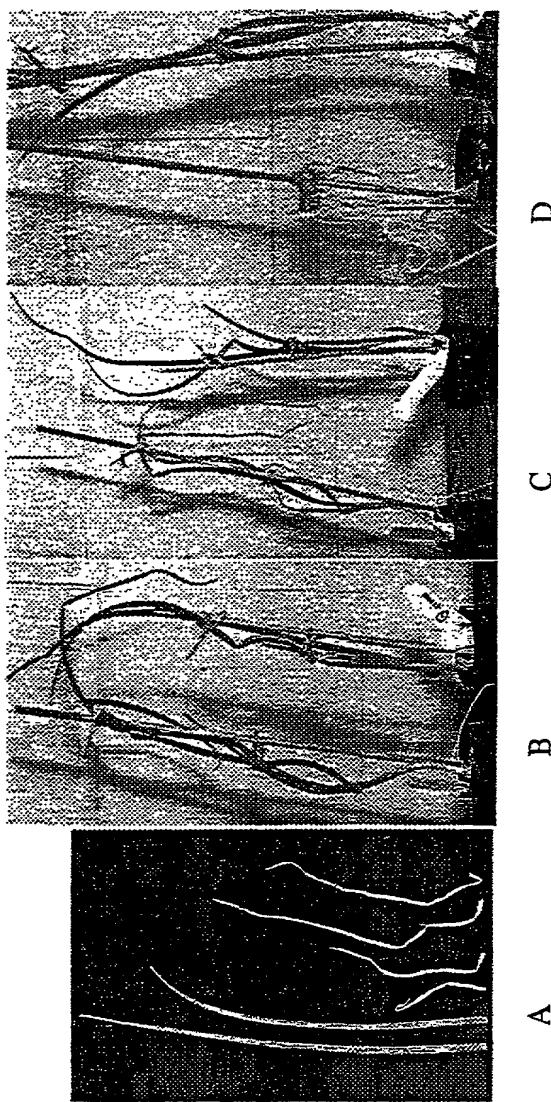


FIGURE 6



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FIGURE 7

DECLARATION AND POWER OF ATTORNEY

As a below named inventor, I hereby declare that: My residence, post office address and citizenship are as stated below next to my name; I believe I am the original, first and sole inventor (if only one name is listed below) or an original, first and joint inventor (if plural names are listed below) of the subject matter which is claimed and for which a patent is sought on the invention entitled:

TRANSFORMATION AND REGENERATION OF ALLIUM PLANTS

the specification of which:

is attached hereto; or

was filed as United States Application Serial No. _____
on July 26, 2001, and was amended on _____
(if applicable); or

was filed as PCT International Application Number PCT/NZ99/00214
on December 10, 1999 and was amended on September 26, 2000
(if applicable).

I hereby state that I have reviewed and understand the contents of the above-identified specification, including the claims, as amended by any amendment referred to above. I acknowledge the duty to disclose information which is material to patentability as defined in 37 CFR § 1.56.

I hereby claim foreign priority benefits under 35 U.S.C. § 119(a)-(d) or § 365(b) of any foreign application(s) for patent or inventor's certificate or § 365(a) of any PCT international application(s), designating at least one country other than the United States, listed below and have also identified below any foreign application(s) for patent or inventor's certificate, or any PCT international application(s) having a filing date before that of the application(s) of which priority is claimed:

Country	Application Number	Date of Filing	Priority Claimed Under 35 U.S.C. 119
New Zealand	333992	January 29, 1999	<input checked="" type="checkbox"/> YES <input type="checkbox"/> NO
			<input type="checkbox"/> YES <input type="checkbox"/> NO

I hereby claim the benefit under 35 U.S.C. § 119(e) of any United States provisional application(s) listed below:

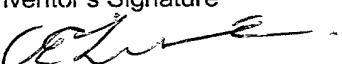
Application Number	Date of Filing

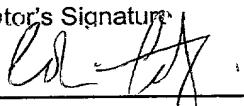
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Application Number	Date of Filing	Status (Patented, Pending, Abandoned)

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